Chem 112L

Department of Chemistry and Biochemistry
University of California, Santa Barbara
Spring 2004

General course information for Chem112L,

BIOPHYSICAL AND BIOANALYTICAL LABORATORY

Lecturer: Kalju Kahn  kalju@chem.ucsb.edu  Phone: x6157
PSB-N 1511
Office hours: Fri 1:00-2:00, "Open Door" policy other times

Teaching assistants:

Section 1:
- Dave Hiller  hiller@chem.ucsb.edu  Phone: x2677  Room: 1142
- Nathan Uter  uter@lifesci.ucsb.edu  Phone: x2677  Room: 1142
Office hours: Tue 3:00-4:00 (Uter); Fri 11:00-12:00 (Hiller)

Section 2:
- Ellie Franzen  efranzen@chem.ucsb.edu  Phone: x2677  Room: 1142
- Richard Chapleau  rchapleau@chem.ucsb.edu  Phone: x4930  Room: 4650
Office hours: Thu 10:00-11:00 (Franzen); Fri 10:00-11:00 (Chapleau)

Lab session times:
Section 1: Monday/Wednesday  2:00-5:50p.m.  PSB-N 2619
Section 2: Tuesday/Thursday  2:00-5:50p.m.  PSB-N 2619

Course website: http://chem.ucsb.edu/~kalju/chem112L


Course Goals
The purpose of Chem 112L is twofold: (i) introduce you to the instrumentation used in biophysical and bioanalytical chemistry, and (ii) help to understand concepts that are taught in Chem112. Your work is organized around six projects, and most projects will take more than one lab session. Lectures and discussions that cover topics that are not normally taught in Chem112 will be held on some days.
Course Information and Expectations of Students:

General
Chem 112L is a laboratory course, in which the main portion of your grade is earned by planning and performing experiments, documenting your work in your lab report, and answering questions in the manual. There is a quiz at the beginning of each experiment, two exams throughout the course, and a poster research presentation at the end of the course. As with any laboratory course, standard lab fee is collected from students who stay beyond the standard drop deadline.

Attendance
Attendance in all laboratory sessions is mandatory. Please contact your instructor, your TA, and your lab partner at least one week ahead of time if you have to miss a class. For each missed class you must turn in a signed letter detailing the reason for missing the class. For unexpected misses, you must provide a verifiable documentation stating why you could not make the class. There is a make-up day toward the end of the quarter to repeat missed or unsuccessful experiments.

Preparation for the lab
Laboratory manuals can be downloaded from the course website: http://www.chem.ucsb.edu/~kalju/chem112L. You are expected to be well familiar with material in the course manual as well as with any material presented in the ‘required reading’ section. You should show an adequate preparation in order to perform the experiment. Your preparation is judged to be inadequate if you receive 33% or fewer points both on your quiz and on your prelab for that day. Your preparation is also judged to be inadequate if you receive 20% or fewer points on either the prelab or the quiz for that day. If you are not adequately prepared, you are asked to leave the lab. You may make up the experiment during a suitable time if this was the first instance of inadequate preparation. You will receive zero points for the whole experiment on any subsequent instance of inadequate preparation.

Academic honesty
Honesty and academic integrity must be always preserved. It is your responsibility to be familiar with common sense about academic integrity. Some examples of academic dishonesty are:

a) altering experimental data without clearly explaining the nature of alterations
b) intentional misrepresentation of the meaning of data
c) using other student’s data as if it was your data
d) copying information from any source and presenting it as if it was your original answer
e) copying images from any source and presenting them as if they were created by you
f) sharing information about quiz questions with other students
g) altering the experimental set-up of other students without their permission

For additional information about types of disallowed conduct, please see http://www.ubalt.edu/studentaffairs/handbook/academic_dishonesty.html

While working with your partner(s) is encouraged in the laboratory, you must write your notebook up independently. You may discuss prelab questions and additional questions with other students in the class but make sure that your answers are original. Plagiarism will not be tolerated and will result in score zero on that lab report. Cheating in any
form will result in a failing grade and notification of Associate Dean of Students, Conduct and Student Relations. No supplemental materials should be used during quizzes and the final exam.

Safety
Even though we have had an excellent safety record in our biochemistry laboratory, the teaching laboratory can be a dangerous place. A few hazards that are present this quarter include hot water or hot surfaces, toxic or corrosive chemicals, electricity, operating centrifuges, and broken glass. The general advice to safety is: **know what you are doing by preparing for lab.** Each experiment in the manual outlines most serious hazards that are present while performing the experiment and discusses ways to prevent accidents. Be sure to read these carefully and ask your TA or the instructor if you have any questions. Your TA will remind you of the potential hazards before each class.

You must follow basic safety rules to ensure safety for yourself and fellow students during the class.

1) Always wear some sort of protective eyewear. You must wear either lab goggles that protect from the sides as well as from the front or a face shield. You can purchase lab goggles during the first weeks of the class from the bookstore. There are four face shields in the laboratory. If you are wearing normal glasses, wear goggles over them as normal glasses do not provide side-protection. You may wear contact lenses along with lab goggles. Protective eyewear is not required in the computer lab.

2) Wear gloves when required, or when you are working with dangerous chemicals. Replace gloves when they become contaminated.

3) Wear appropriate clothing in the wet lab. You must wear closed shoes, and a shirt that covers the midsection. No shorts are allowed, and long pants and long sleeves are recommended.

4) Label all solutions that you prepare clearly. If the solution is in a container larger than 5 mL, make sure that the label conveys information about the content of the container and also identify your group or you personally as the person responsible for this solution. Unlabeled solutions will be discarded.

5) Familiarize yourself with the safety equipment in the lab. Our lab has one fire extinguisher near the back door, and eyewash station near the front door. The first aid kit is in the top right drawer next to the main sink. Emergency phone numbers are posted in the lab. In case of fire, earthquakes, or other major disasters leave the lab if this is safe, and meet your instructor at the lawn south of PSB-N building.

6) Do not eat or drink in the laboratory. Closed containers with food may be brought into the laboratory but must be stored in your bag or backpack placed in the cabinet box. You may eat or drink outside the laboratory if time permits.

7) Never work alone. Never use mouth suction. Never open the lid of a spinning centrifuge. Do not perform any unauthorized experiments. Come and see your instructor if you would like to perform an additional interesting or fun experiment. He is probably willing to let you do it.

8) In case of accident, alert fellow students and immediately take an appropriate action. Explain to your TA what happened and seek further help if necessary.

Ignoring safety rules while in the laboratory will lead to oral warning on the first instance, and deduction of lab quality points for each following instance. Serious intentional violations of safety rules will lead to the dismissal from the course.
Cleanliness
It is important to maintain cleanliness in this laboratory. Even minor impurities on the glassware or on the pipette tip may ruin an otherwise well-done biochemical experiment. For example, using the same pipette tip to transfer two enzymes from their containers into your microcentrifuge tube will most surely contaminate the stock of the second enzyme with the first one and will likely ruin the results for the whole class. You will be working a lot with pipettors that use disposable tips. Discard the tip as soon as you do not need it. They are a lot cheaper than the chemicals that you are working with.

Most used plasticware, such as microcentrifuge tubes or Falcon tubes (15 mL, 50 mL size) are for one-time use. Empty all the tubes before discarding them (it’s OK to leave less than 50 µL in microcentrifuge tubes). Discard broken glassware and used glass pipettes into the red container. Do not discard functional glassware or any parts of the equipment used. If your glassware is visibly dirty, wash it with soap and hot water, otherwise rinse several times with distilled water from the tap, and place on the drying racks. Do not leave any dishes in the sink.

Student must label all the containers larger than 5 mL that contain solutions made by them clearly with their name and content of the container. Unlabeled containers will be disposed by your instructor. Each student is responsible for completely cleaning the workplace, washing all the glassware used, and disposing all the disposable plastic-ware. Your instructor will adjust your grade based on your work ethics. Leaving a mess will result in a bad grade.

Experiments and lab reports
There are six major projects and possibly one or two instrument demonstrations. The six projects are:
1. Allantoin conformational analysis: computer modeling and 2D NMR
2. Protein unfolding monitored by circular dichroism (unfolding of lysozyme)
3. Ligand binding by lysozyme: determination of the equilibrium constant
4. Electrospray ionization mass spectrometry characterization of proteins
5. Enzyme inhibition kinetics: glyceraldehyde-3-phosphate dehydrogenase
6. Protein crystallography

Each report must be typed and look professional as if it was a manuscript that you submit for a publication. Each report should include the following (percentage of grade shown in parentheses):
- title page giving the name of the experiment, your name, and grading rubric
- the objective of the experiment in your own words (3 %)
- the rationale for the experimental set-up (10 %)
- methods: a concise summary of steps taken to accomplish the goal. Describe sample preparation. (2-3 %)
- answers to the prelab questions (17 %)
- most important data organized as tables, graphs, or images (10 %)
- mathematical analysis of data, if applicable (7-8 %)
- a discussion about the physical meaning of your results (20 %)
- a conclusions section summarizing the most important findings (10 %)
- answers to the additional questions given at the end of the experiment, (20 %)
- an appendix with all the raw data that were collected, and all calculations that you performed
The objective of the experiment, the rationale for the experimental set-up, concise summary of the procedure, and answers to your prelab questions constitute the prelab that you must prepare before performing each experiment. The rationale for the experimental setup is very important concept that many students find confusing. In this section, you should explain how the methods that you are going to use in the class allow you to achieve the objective of the lab. Typically, you should show how the technique you will use allows study of the property of interest in your system. You should also offer a plan to interpret your data and predict at least one reasonable outcome. Please note that this section is not a copy of your lab manual’s background section nor a place to repeat detailed experimental procedures but involves thoughtful synthesis of all the information available to you.

Prelab questions
The prelab questions are designed to help you to understand the experiment that you are doing. Answer your prelab questions directly into your notebook but make an additional carbon copy (or photocopy) of the answers and bring the copy with you to the class. Your TA will check your prelab and collect the carbon copy with your answers to the prelab questions. He will also tell you if any of your answers to prelab questions were wrong. You may correct your wrong answers and get up to 25% of credit on the wrong answer when you turn your notebook in for final grading. Please mark clearly the corrected answer. Answers to the prelab questions typically contribute about 14% of your total lab report points.

Experiments and group work
You will perform series of experiments in this laboratory and complete several computer assignments. Because of the large number of students and limited resources, students will work in groups to perform the experiments. Computer assignments will be performed independently. In order to best use the available time, all the students in one section are assigned into one of the four groups. The schedule at the end of this syllabus outlines experiments that each group does on a particular day.

Quizzes
There will be six quizzes in this lab on the first day of a related experiment. There is no quiz on the first day of the class; the allantoin conformational analysis quiz will be given at the time of NMR experiment. All quiz scores count, so the maximum number of quiz points is 60.

Exams
There will be two exams. The first will be on protein folding by CD measurements, mass spectrometry, and ligand binding equilibria. The second will cover enzyme kinetics, protein crystallization, and structure determination by NMR. The purpose of the exams is to evaluate (1) your understanding of the theory behind the technique and (2) your ability to analyze the physical meaning of the data obtained. Each exam is worth 50 points, and the total maximum from two exams is 100.

Poster presentation
There will be a poster presentation at the end of the quarter. Currently, the poster session is planned for noon to 2 PM for Thursday, June 3. The poster presentation is worth 20 points.
Grading

Your grade will be based on the number of points you earn out of 450 points possible. There are six lab write-ups. You may make up failed experiments during the last two weeks. The points for each experiment are as follows:

- Allantoin conformational analysis: computer modeling and 2D NMR 50
- Protein unfolding monitored by circular dichroism (unfolding of lysozyme) 40
- Ligand binding by lysozyme: determination of the equilibrium constant 40
- Electrospray ionization mass spectrometry characterization of proteins 40
- Enzyme inhibition kinetics: glyceraldehyde-3-phosphate dehydrogenase 40
- Protein crystallography 40
- Lab work ethics/cleanliness 20

**Total Lab** 270

- Exams 100
- Quizzes 60
- Poster presentation 20

**Total Theory** 180

**Grand Total** 450

Last year, students who collected over 82% of total possible points received grade A- or better. The percentage of students who received A- or better was 26%. Last year, students who collected less than 60% of total possible points received grade C- or lower. The percentage of students who have received grade C- or lower in this course has been around 15% in past.

_Wishing you the best success in this course_  
-- Kalju --